

ISOLATION AND IDENTIFICATION OF FUNGI ASSOCIATED WITH LEAF BLIGHT DISEASE OF MISTLETOE (*TAPINANTHUS* SP.)



OBANS, U. H¹. AND YAHAYA A^{1*}

Department of Botany, Faculty of Life Sciences, Ahmadu Bello University, Zaria, Nigeria. *Corresponding Author: <u>ayahaya@abu.edu.ng</u>

Received: February 14, 2025, Accepted: April 28, 2025

Abstract:	Mistletoes (<i>Tapinanthus</i> sp.) are hemi-parasitic flowering plants that derive water and minerals from their heat elected after resulting in significant physiclesical impacts. This study investigated the fungel appairs
	nost plants, other resulting in significant physiological impacts. This study investigated the fulligat species
	associated with leaf blight in mistietoe leaves. Symptomatic and asymptomatic leaves were collected from
	Terminalia catappa host trees at Ahmadu Bello University, Zaria, Nigeria, and analyzed in the Mycology
	Laboratory, Department of Botany, Ahmadu Bello University, Zaria, using 39g of Potato Dextrose Agar
	(PDA) prepared in 1000 ml of distilled water. Six fungal species were isolated and identified: Aspergillus sp.
	Gloeosporium sp. Rhizoctonia sp. Cunninghamella sp. Sclerotium sp. and Phoma sp. Among these,
	Aspergillus sp. was the most frequently occurring species, appearing in 33.3% of symptomatic leaves and
	55.5% of asymptomatic leaves. This study highlights the diversity of fungi associated with mistletoe leaf
	blight and underscores their potential ecological and pathological significance.

Keywords: Fungal pathogens, Isolation, Identification, leaf blight, mistletoe, parasitic plant

Introduction

Mistletoes (Tapinanthus sp.) are hemi-parasitic plants (Moreira, et al., 2019) commonly found in tropical and subtropical regions which belongs to the class Magnoliopsida, subclass Rosidae, order Santales (Judd et al., 2002). They depend on their host plants for water and minerals, which often leads to salt imbalances and physiological stress in the host (Milius, 2000). The two major families of mistletoe are Loranthaceae and Viscaceae. Most genera of African mistletoe belong to the family Loranthaceae (Dambele et al., 1994). It is the largest family of mistletoe having up to 73 genera and over 900 species (Judd et al., 2002). The six major genera of Loranthaceae found in Nigeria are Tapinanthus, Agelanthus, Loranthus, Globometula, Phragmenthera and Englerina, with Tapinanthus being far more widespread in the Nigerian savanna (Bako et al., 2001). Mistletoes are parasitic plants that divert water and use far more water per unit carbon fixed during photosynthesis than their hosts (Scalon and Wright, 2015). All mistletoes produce a morphologically diverse structures (haustoria) which allow them to interface with their host (Calvin and Wilson, 2006). Trees can respond to mistletoes infection by producing sclerophyllous, yet less productive leaves (Scalon, et al., 2017). According to studies conducted over the past few years, the intensity of mistletoe damage on trees in countries increases from year to year (Varga, et al., 2012).

Seeds of most mistletoe are spread by birds that eat the fruits (Cowles, 1964) or by wind. Because of their easy dispersal and widespread tropism, mistletoes have become serious pests for commercial fruit and timber plantations. Mistletoes have economic importance as medicinal plants (Szurpnicka, *et al.*, 2020), but their parasitic nature poses challenges to forestry and agriculture (Mudgal *et al.*, 2022). Boussium *et al.* (2004) reported that mistletoe (*T. globiferus*) parasitized 126 species. Infection by mistletoe leads to water and nutrient stress of the host plant and deteriorates its healthy establishment and survival. Infection by mistletoes also triggers host plant responses, varying from mechanical to chemical mechanisms which

are analogous to herbivory defences, and negatively impacts host plant growth and reproduction. (Muche *et al.*, 2022).

Approximately 40 % of moderate and high level infestation rates increase the possibility of tree mortality in the absence of management applications (Zúñiga *et al.*, 2024).

In contrast, ecologists may see mistletoe as a friend, in light of the wildlife habitat, biodiversity and nutrient cycling it p romotes (Griebel *et al.*, 2017).

Leaf blight is a major fungal disease that affects mistletoe, causing necrotic lesions and reducing the plant's photosynthetic capacity. Although mistletoes have been studied extensively from botanical and ecological perspectives, the fungal communities associated with leaf blight remain poorly understood (Mayer, 2006).

This study aims to address this knowledge gap by isolating and identifying fungi associated with leaf blight in *Tapinanthus* sp. The findings could have implications for biological control strategies and the management of fungal diseases in economically significant host plants.

Materials and Methods

Study Area

The study was conducted in the Mycology Laboratory of the Department of Botany, Ahmadu Bello University, Zaria, Kaduna State, Nigeria.

Sample collection

Symptomatic and asymptomatic mistletoe leaves were collected from *Terminalia catappa* trees within the university campus. The collected leaves were transported to the laboratory in sterile polyethylene bags for further analysis.

Preparation of PDA and Fungal Isolation

Potato Dextrose Agar (PDA) was prepared by dissolving 39 g of PDA in 1,000 ml of distilled water and sterilized at 121°C for 15 minutes. Chloramphenicol (500 mg) was added to inhibit bacterial growth (Deep and Emmanuel, 2015)

Symptomatic leaf tissues were surface-sterilized with 1% sodium hypochlorite for 3 minutes and then rinsed in three changes of sterilized distilled water, and plated on PDA. Plates were incubated at room temperature (25–28°C) for seven days (Okigbo and Osuinde, 2003) and pure cultures were obtained through sub-culturing.

Identification of Fungal pathogen

Fungal identification was based on macroscopic (colony morphology) and microscopic (conidial, spore, and hyphal structures) features. Slides were prepared using lactophenol cotton blue stain and examined under a light microscope at $10 \times$ and $40 \times$ magnification. Identifications were confirmed using standard fungal manuals.

Results

Six fungal species were isolated from symptomatic and asymptomatic mistletoe leaves: *Aspergillus* sp. (Plate II and III) *Rhizoctonia* sp. (Plate IV), *Cunninghamella* sp. (Plate V), *Gloeosporium* sp. (Plate VI) *Sclerotium* sp. (Plate VII) and *Phoma* sp. (Plate VIII). The percentage occurrence of each species is presented in Figure 1 and 2. *Aspergillus* sp. was the most frequently occurring fungus, found in both symptomatic (33.3%) and asymptomatic leaves (55.5%).





Plate 1. A – Mistletoe leaf blight with light brown lesions becoming necrotic used for isolation B. Asymptomatic mistletoe leaf from same plant



Plate II: Pure culture of Aspergillus flavus, A. Macroscopic view,



B. Microscopic view of Aspergillus flavus





Plate III: Pure culture of Aspergillus niger

A. Macroscopic view,

B. Microscopic view of Aspergillus niger





Plate IV: Pure culture of Rhizoctonia sp.

A. Macroscopic view

B. Microscopic view of Rhizoctonia sp.



Plate V: Pure culture of Cunninghamella sp. A. Macroscopic view, B. Microscopic view of Cunninghamella sp.





Plate VI: Pure culture of Gloeosporium sp.

A. Macroscopic view

B. Microscopic view of Gloeosporium sp.



Plate VII: Pure culture of Sclerotium sp.



A. Macroscopic view,

B: Microscopic view of Sclerotium sp.





Plate VIII: Pure culture of Phoma sp. A. Macroscopic view b. Microscopic view of Phoma sp.



Fig 1: Percentage occurrence of Isolated Fungi Pathogens from symptomatic Mistletoe leave.



Fig 2: Percentage occurrence of Isolated Fungi Pathogens from asymptomatic mistletoe

Discussion

The study revealed that, nine fungi were isolated from infected leaves and nine from uninfected leaves which comprises 6 different fungi species, Rhizoctonia sp., Cunninghamella sp., Phoma sp., Aspergillus sp., Sclerotium sp. and Gloeosporium sp. Out of the 6 fungi species, Aspergillus sp. was found to be the most frequently encountered from both asymptomatic and symptomatic leaves consisting of 8 fungi isolates with 33.3% from symptomatic leaves and 55.5% from asymptomatic leaves. Aspergillus sp. is the predominant fungal pathogen associated with mistletoe leaf blight. Aspergillus sp. has been reported as one of the causal agent of blight on mistletoe leaves by (Kotan et al., 2012). Aspergillus is a genus of fungi that consists of several hundred species. Some which are significant human pathogens such as A. flavus which can cause aspergillosis. A. niger is also commonly known for producing black mold on fruits and vegetables (Samson et al., 2014). Latiffah, (2024) reported that the genus Aspergillus contains several species that are important plant pathogens. Pawłowicz et al., (2024) also reported four though different fungi as a saprotroph or parasite on the leaves and twigs of mistletoe in Poland. Rhizoctonia sp was also isolated in this study. Similarly, Rhizoctonia solani reported as a soil-borne fungal pathogen that causes disease in a wide range of plants worldwide (gonzalez et al., 2011). Rhizoctonia solani is an important plant pathogen that causes yield losses in legume crops worldwide, causing severe diseases such as blights. damping-off, and rots (Basbagci and Dolr, 2020). Gloeosporium sp isolated from this study was frequently

isolated from blighted shoot and leaves of young plantation trees and nursery seedlings of Eucalyptus nitens and E. globulus in Australia (Yuan et al., 2000). Similarly. species of Gloeosporium, *G.* (*Colletotrichum*) gloeosporioides, was

reported to cause anthracnose disease of vegetables and fruits, it lives saprophytically on crop residues, attacking the tissues at the crown and base of the stems of more developed plants (Cho et al., 2003). However, Schaible et al., (2015) isolated Gloeosporium sp as an endophyte of Tsuga heterophylla and that the volatile phase of this endophyte was active against a number of plant pathogenic fungi including Phytophthora palmivora, Rhizoctonia solani, Ceratocystis ulmi, Botrytis cinerea, and Verticillium dahlia. Sclerotium sp was one of the fungi isolated in this study. Similarly, Le et al., (2012) reported Sclerotium rolfsii as causative agent of white rot and one of the destructive pathogens of plants worldwide. Phoma sp isolated in this study has also been reported as a severe field pathogen of onion (Badillo-Vargas et al., 2008; Boerema et al., 2004)

The presence of fungi in asymptomatic leaves suggests that some maybe endophytic or latent pathogens. Other species, such as Rhizoctonia sp. and Gloeosporium sp. were also significant, causing necrotic lesions on leaves. These findings emphasize the need for further research on the ecological roles of these fungi.

Conclusion

The study isolated and identified Aspergillus sp. Rhizoctonia sp. Sclerotium sp. Phoma sp. Gloeosporium sp. and *Cunninghamella* sp. from symptomatic and asymptomatic leaves from same mistletoe plants.

Aknowledgements

Special thanks go to Ms. Ramatu of mycology laboratory, Department of Botany, Ahmadu Bello University, Zaria for her guide throughout the practical process.

References

- Bako, S. P., Onwuchekwa, B. N., Bako, L. S. P. and Iortsum, D. N. (2001). Physiology of the African Mistletoe (*Tapinanthus dodeneifolius* (DC) DANSER) and its influence on the metabolism of two hosts (*Albizia lebbeck* Benth and *Citrus sinensis* L.) in Nigeria. *Journal of Agriculture and Environment* 2(10): 81-92.
- Badillo-Vargas E. I., Rivera-Vargas L. I., and Calle-Bellido
 J. (2008). Morphological, pathogenic and molecular characterization of Phoma spp. Isolated from onion field soils in Puerto Rico. The Journal of
- Boerema, G. H., J. de Gruyter, M. E. Noordeloos and M. E. C. Hamers, (2004). Phoma Identification Manual: Differentiation of Specific and Infra-specific Taxa in Culture.CABI Publishing. Wallingford, Oxfordshire, UK.
- Boussim IJ, Guinko S, Tuquet C, Salle G (2004). Mistletoes of the agroforestry parklands of Burkina Faso. Agrofor. Syst. 60:39-49.
- Calvin, C. L., and Wilson C. A. (2006). "Comparative Morphology of Epicortical Roots in Old and New Loranthaceae with Reference to Root Types, Origin, Patterns of Longitudinal Extension and Potential for Clonal Growth." Flora 201:51-64.
- Cho S., Sam Lee., Jin Cha B., Kim Y. H., Shin K. (2003). Detection and characterization of the *Gloeosporium* gloeosporioides growth inhibitory compound iturin A from Bacillus subtilis strain KS03. FEMS Microbiology Letters, 223(1) P.47 –51. ISSN 0378-1097, https://doi.org/10.1016/S0378-1097(03)00329-X.
- Cowles RB (1964). Field notes of a naturalist in South Africa. University of California Press, California, USA. P 114.
- Dambele, D. A. Raynal-Rouques, G. and Tunguer, S. (1994). Parasitic plants of crops and trees in the sahel. Sahel Institute 432pp.
- Deep, A. and Emanuel, W. (2015). *Laboratory Guide for General Microbiology*. University of Califonia. 5-17.
- Gonzalez, M., Pujol, M., Metraux, J.-P., Gonzalez-Garcia, V., Bolton, M.D. And Borrás-HIDALGO, O. (2011), Tobacco leaf spot and root rot caused by *Rhizoctonia solani* Kühn. Molecular Plant Pathology, 12: 209-216. <u>https://doi.org/10.1111/j.1364-3703.2010.00664.x</u>
- Griebel A., Watson D., Pendall E. (2017). Mistletoe, friend and foe: Synthesizing ecosystem implications of mistletoe infection. Environmental Research Letter 12(11). DOI <u>10.1088/1748-9326/aa8fff</u>
- Judd, W. S., Campbell, C. S., Kellogg, E. A., Stevens, P. F. and Donahue, M. J. (2002). *Plant systematics*.
- Kotan, R., Okutucu, A., Gormez A. A., (2012). (Potential Application in Biocontrol. Journal of Phytopathology 161(3) DOI: 10.1111/jph.12048
- Latiffah Z. (2024). An Overview of *Aspergillus* Species Associated with Plant Diseases. Pathogens. 2024 Sep 20;13(9):813. doi: 10.3390/pathogens13090813. PMID: 39339004; PMCID: PMC11435247.
- Le C.N., Kruijt M., Raaijmakers J.M. (2012). Involvement of phenazines and lipopeptides in interactions between Pseudomonas species and Scler rolfsii, causal agent of stem rot disease on groundnut. Journal of Applied Microbiology. 112(2):390–403. doi: 10.1111/j.1365-2672.2011.05205.x.
- Mayer AM. (2006). Pathogenesis by fungi and by parasitic

plants: similarities and differences. *Phytoparasitica* **34**: 3–6.

Milius S (2000). Botany under the mistletoe. Science News, 158:412

Moreira F. A., Costa L.T., Ceccantini G., Furlan C. M. (2019). Mistletoe effects on the host tree Tapirira guianensis: insights from primary and secondary metabolites. Chemoecology 29(1). DOI: 10.1007/s00049-018-0272-6

- Muche, M., Muasya, A.M. and Tsegay, B.A.(2022) Biology and resource Responses of host plants. *Ecol Process* **11**, 24 .<u>https://doi.org/10.1186/s1371</u> 7-021-00355-9
- Mudgal G, Kaur J, Chand K, Parashar M, Dhar SK, Singh GB, Gururani MA (2022) Mitigating the Mistletoe Menace: Biotechnological and Smart Management Approaches. Biology (Basel). 11(11):1645 doi: 10.3390/biology11111645.
- Okigbo, R., and Osuinde, M. (2003). Fungal leaf spot Diseases of Mango (Mangifera indica L.) in South-eastern Nigeria and Biologicl Control with Bacillus subtilis. Okigbo, R. N and Osuinde, M. I. Fungal leaf Spot Diseases of Mango (Mangifera indicplant protect. Sci, 39(2):70-77.
- Pawłowicz, T., Oszako, T., Borowik, P., Malewski, T., & Faedda, R. (2024). The Mycobiota Diversity Associated with Mistletoe (*Viscum album* L.) in Poland. *Forests*, 15(12), 2152. https://doi.org/10.3390/f15122152
- Samson, R. A., Visagie, C. M., Houbraken, J., Hong, S. B., Hubka, V., Klaassen, C. H. W., et al (2014). Phylogeny, identification and nomenclature of the genus Aspergillus. Stud. Mycol. 78, 141–173.doi: 10.1016/j.simyco.2014.07.004
- Scalon M. C., and Ian Wright J. (2015). A global analysis of water and nitrogen relationships between mistletoes and their hosts: broad-scale tests of old and enduring hypotheses. Functional Ecology 29 (9) (2015), pp. 1114-1124. doi: 10.1111/1365-2435.12418
- Scalon MC, Wright IJ (2017) Leaf trait adaptations of xylemtapping mistletoes and their hosts in sites of contrasting aridity. Plant Soil 415:117–130. https://doi.org/10.1007/s11104-016-3151-3
- Schaible GA, Strobel GA, Mends MT, Geary B, Sears J (2015). Characterization of an Endophytic Gloeosporium sp. and Its Novel Bioactivity with "Synergistans". Microbial Ecology.70(1):41-50. doi: 10.1007/s00248-014-0542-y. Epub 2014 Dec 11. PMID: 25501886.
- Szurpnicka A, Kowalczuk A, Szterk A. (2020). Biological activity of mistletoe: in vitro and in vivo studies and mechanisms of action. Arch Pharm Res. Jun;43(6):593-629. doi: 10.1007/s12272-020-01247-w. Epub PMID: 32621089; PMCID:PMC7340679.
- Varga I, Taller J, Baltazar T, Hyvonen J, Poczai P (2012). Leaf-spot disease on European mistletoe (Viscum album) caused by Phaeobotryosphae-ria visci: a potential candidate for biological control. Biotechnol Lett34:1059–1065
- Yuan, Z. Q., Wardlaw, T., and Mohammed, C. (2000). First report of *Gloeosporidina* sp. isolated from lesions on shoots and leaves of Eucalyptus nitens and E. globulus in Australia. Plant Diseae 84(5): 510-512
- Zúñiga J. J. A., Bobadilla R. L. H., Maass S. F., Fredericksen T. S., Agramont A. R. E. (2024). Occurrence and effect of dwarf mistletoe (Arceuthobium globosum and A. vaginatum) in high-elevation forests in México. Trees, Forests and People, 18 (100706). https://doi.org/10.1016/j.tfp.2024.100706